

## Processing of the Human Heparanase Precursor and Evidence That the Active Enzyme Is a Heterodimer\*

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Human platelet heparanase has been purified to homogeneity and shown to consist of two, non-covalently associated polypeptide chains of molecular masses 50 and 8 kDa. Protein sequencing provided the basis for determination of the full-length cDNA for this novel protein. Based upon this information and results from protein analysis and mass spectrometry, we propose a scheme to define the structural organization of heparanase in relation to its precursor forms, proheparanase and pre-proheparanase. The 8- and 50-kDa chains which make up the active enzyme reside, respectively, at the NH<sub>2</sub>- and COOH-terminal regions of the inactive precursor, proheparanase. The heparanase heterodimer is produced by excision and loss of an internal linking segment. This paper is the first to suggest that human heparanase is a two-chain enzyme.

A vast literature attests to the importance of heparan sulfate proteoglycans (HSPG)<sup>1</sup> in a variety of physiological processes (cf. Refs. 1–8 for reviews and additional references). Not only do these complex molecules provide a physical barrier to movement of cells into tissues, but the carbohydrate, or glycosylaminoglycan moieties of the HSPG bind and sequester a variety of bioactive proteins, including growth factors, chemokines, cytokines, and enzymes. These several proteins may be retained in complexation with the proteoglycans, or released when the HSPG are broken down, thus providing mechanisms for induction of growth, chemotaxis, and extravasation of a diverse set of cells in normal or disease processes (3, 8). This regulatory aspect of HSPG function impacts a wide spectrum of biological phenomena underlying inflammatory and cardiovascular dis-

eases and cancer.

Heparanases, enzymes secreted by activated platelets (2) and neutrophils (9) and by metastatic cells (3), hydrolyze the glycosylaminoglycan units of HSPG, thus facilitating release of the many protein modulators of cell function that are bound at these sites. This helps pave the way for migration of neutrophils and cancer cells from the vasculature into tissues and promotes supply of blood vessels to a growing tumor (3). Although the importance of heparanase function has long been recognized (3), and bacterial heparanases have been thoroughly characterized (10), the mammalian enzyme has remained, until very recently, an elusive target. Now, for the first time, Vlodavsky *et al.* (11), Hulett *et al.* (12), and Kussie *et al.* (13) have reported the cDNA and derived amino acid sequences of a novel, human heparanase. The molecule is unique among known protein structures and shows no homology to the bacterial enzymes. All three groups agree that the heparanase is a single-chain glycoprotein, roughly 50,000 in molecular weight, that is derived from a precursor of 543 amino acids as defined by the full-length cDNA (11–13). Clearly, this important discovery could have major therapeutic implications for the development of new classes of drugs for cancer, heart disease, and inflammation (3, 14, 15).

The present paper describes the purification and characterization of heparanase from human platelets. During the course of this work, we also determined the cDNA and derived protein sequences for heparanase and its precursor molecules, and our results are in complete accord with those from the aforementioned studies. However, in contrast to the conclusion that heparanase is a single 50-kDa polypeptide (11–13), our findings suggest that the active enzyme is a heterodimer in which this 50-kDa protein works in concert with a tightly associated, but non-covalently linked, 8-kDa peptide. Both chains arise by processing of a single precursor protein, designated herein as proheparanase. A scheme is proposed to define the precursor forms of heparanase and the processing events leading to enzyme activation.

### EXPERIMENTAL PROCEDURES

#### Materials

Fresh human platelet concentrates were obtained by apheresis. Complete protease inhibitor tablets and diisopropyl fluorophosphate were purchased from Roche Molecular Biochemicals. H[<sup>35</sup>S]PG substrate was prepared as described by Ledbetter *et al.* (16). Heparin-Sepharose CL6B and Protein A-Sepharose Fast Flow bulk media, and Superdex-75 Hi-Load, and heparin Hi-Trap columns were purchased from Amersham Pharmacia Biotech. Precast 10–20% gradient and 10% homogeneous polyacrylamide gels, Tricine running buffer, and molecular weight markers were obtained from ISS Enprotech. Ultra-free-MC nominal molecular weight limit filter units were obtained from Millipore Corp. Polyvinylidene difluoride membranes were from Schleicher and Schuell. Jupiter C4 and C18 microbore columns were obtained from Phenomenex, and HPLC-grade water and acetonitrile were from OmniSolv. Pyroglutamate aminopeptidase was from Takara Biomedicals.

#### Methods

##### Assay for Heparanase Activity

Samples containing heparanase activity were identified as described earlier (7) by their ability to degrade metabolically radiolabeled and purified high molecular weight H[<sup>35</sup>S]PG derived from mice bearing the Engelbreth-Holm-Swarm tumor (16) to filterable fragments. We define a unit of activity as that amount of enzyme that produces breakdown of 1% of the substrate per hour, as measured by liberation of counts that pass through a 30,000 nominal molecular weight limit membrane filter.

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<sup>1</sup> The abbreviations used are: HSPG, heparan sulfate proteoglycan; Tricine, N-[2-hydroxy-1,1-bis(hydroxymethyl)ethyl]glycine; GSH, glutathione; DTT, dithiothreitol; SEC, size exclusion chromatography; CV, column volume; PAGE, polyacrylamide gel electrophoresis; MALDI-TOF MS, matrix-assisted laser desorption time of flight mass spectrometry; RP-HPLC, reverse phase high performance liquid chromatography; TFA, trifluoroacetic acid.

## Purification of the Platelet Heparanase

**Heparin-Sepharose Chromatography**—The supernatant from activated, centrifuged platelet lysates (7) was made 1 mM in glutathione (GSH) and dithiothreitol (DTT) and loaded (1.0 ml/min) onto a column of heparin-Sepharose (1.6 × 20 cm; 40 ml) equilibrated with phosphate-buffered saline, 1 mM in both GSH and DTT. The column was then washed with 0.01 M sodium acetate, pH 5.0, containing 1 mM GSH, 1 mM DTT, and 0.35 M NaCl (buffer A) until the initial base line was reestablished. At this time, a 750-ml linear gradient of increasing NaCl concentration (0.35–1.5 M) in buffer A was applied to elute the active heparanase. The column effluent was monitored by UV detection at 280 nm, and aliquots were taken for heparanase assay. Fractions (9 ml) containing active heparanase were pooled and concentrated using a stirred cell ultrafiltration module.

**Superdex-75 Size Exclusion Chromatography**—The ultrafiltration retentate (typically 8–10 ml) was loaded in 1-ml batches directly onto a Superdex-75 Hi-Load size exclusion chromatography (SEC) column (1.6 × 60 cm) equilibrated with 0.01 M sodium acetate, pH 5.0, containing 0.5 M NaCl, 1 mM DTT, and 10 mM  $\beta$ -octyl glucoside (buffer B). The column was run at 1.0 ml/min, and the effluent was monitored at 280 nm. Fractions of 4–6 ml were collected, and aliquots were assayed by SDS-PAGE  $\pm$  Western blotting and for heparanase activity.

**Heparin Hi-Trap Chromatography**—Pooled fractions from the SEC step were diluted 2-fold with deionized water to lower the ionic strength and loaded directly onto a 1-ml column of heparin Hi-Trap-Sepharose equilibrated with 0.01 M sodium phosphate, pH 7.0, containing 0.25 M NaCl, 1 mM DTT, and 10 mM  $\beta$ -octyl glucoside (buffer C). The column was run at 0.5 ml/min and the effluent monitored at 280 nm. Once the original base line was reestablished, the column was washed with buffer C for an additional 5 column volumes (CV), followed by a biphasic gradient of increasing NaCl concentration (5 CV of 0.25–0.63 M NaCl in buffer C followed by 23 CV of 0.63–1.5 M NaCl in buffer C). Fractions of 2 ml were collected, and aliquots were assayed for activity and by SDS-PAGE  $\pm$  Western blotting.

## Protein Characterization

**SDS-PAGE**—SDS-PAGE (17) was performed under both reducing and non-reducing conditions employing the Tricine buffer system (18) and 10–20% gradient polyacrylamide gels (10 × 10 cm). After electrophoresis, gels were either stained with silver (19) or transferred to polyvinylidene difluoride membranes for Western blot analysis.

**Protein Sequence Analysis and Mass Spectrometry**—Peptide and protein sequences were obtained by automated Edman degradation in a PE-Biosystems Procise Model 494 protein sequencer. Model 610A Version 2.1 software was employed for data acquisition and processing. Matrix-assisted laser desorption time of flight mass spectroscopy (MALDI-TOF MS) was performed on a Perseptive Biosystems Voyager Elite mass spectrometer.

## Antibody Preparation and Immunological Methods

After the cDNA and derived protein sequences for heparanase were in hand, antibodies were raised against peptide antigens taken from various regions of the enzyme sequence. The following synthetic peptides were prepared by conventional solid phase methods:  $\text{NH}_2$ -R<sup>273</sup>KTAKMLKSF<sup>283</sup>LKAGGEVIGGC-COOH and  $\text{NH}_2$ -V<sup>399</sup>WLGETSSAYGGC-COOH, from the 50-kDa subunit, and  $\text{NH}_2$ -D<sup>108</sup>PKKESTFEERSGGC-COOH, overlapping the COOH terminus of the 8-kDa peptide and the segment linking the 8- and 50-kDa peptides. These peptides were conjugated to a maleimide-activated form of keyhole limpet hemocyanin, and the keyhole limpet hemocyanin-conjugated peptides were injected into rabbits. Verification that the antisera specifically recognized the peptides, proheparanase, and the 8- and 50-kDa polypeptides of active platelet heparanase was accomplished by Western blotting.

## RESULTS AND DISCUSSION

**Purification of Platelet Heparanase**—Chromatography of the crude platelet lysate on a column of heparin-Sepharose yielded a profile similar to that published earlier (7). Elution of the column with a gradient of increasing [NaCl] led to recovery of all of the heparanase activity in a region from about 0.8–1.2 M NaCl. SDS-PAGE analysis of the resulting fractions revealed no hint of the heparanase on stained gels, but Western blot analysis using antibodies against heparanase peptide segments correlated the presence of a 50-kDa band with fractions

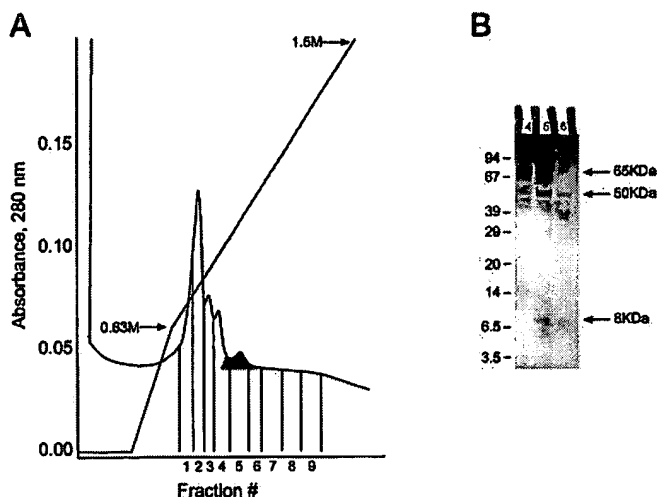


FIG. 1. Heparin Hi-Trap chromatography of the heparanase fractions obtained from size exclusion chromatography. A, elution profile; activity measurements revealed that fraction 5 (shaded) contained most of the heparanase activity, with lesser amounts in fractions 4 and 6. B, SDS-PAGE of fractions 4–6 under non-reducing conditions. Two protein bands of masses approximately 8 and 50 kDa are clearly evident by silver staining in the active fraction (lane 5). Western blotting of an identical gel using peptide antibodies to heparanase showed that both bands are recognized, as well as the 65-kDa proheparanase band seen in fraction 4. All pertinent proteins are designated with arrows.

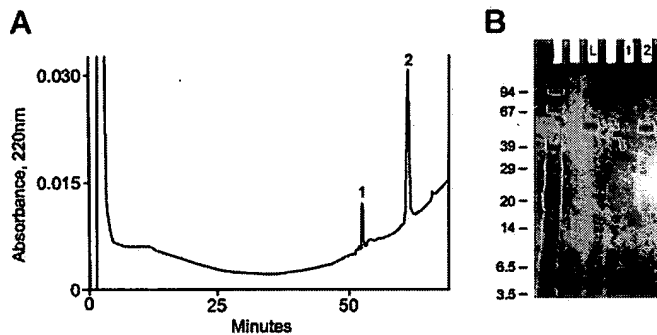


FIG. 2. RP-HPLC of purified platelet heparanase. A, proteins were resolved on a Jupiter C4 reverse-phase column developed over 70 min with a linear gradient of increasing acetonitrile concentration from 0 to 70% in 0.15% TFA. B, SDS-PAGE of the RP-HPLC peaks shows that peak 1 correlates with the 8-kDa (lane 1) and peak 2 with the 50-kDa chain (lane 2) identified in the load (L). The ratio of areas under the peaks suggested a near molar equivalence of the 8- and 50-kDa species.

having heparan sulfate-degrading activity. A 65-kDa band was also observed by Western blotting of this fraction. The second purification step by size exclusion chromatography gave high resolution of heparanase activity in a peak comprising components of about 20–50 kDa as seen on SDS-PAGE. This peak was clearly resolved from the elution position of chemokines (7).

The pool of heparanase activity from the SEC step was loaded onto the final heparin Hi-Trap affinity column, and the elution profile is shown in Fig. 1A. A small peak, shaded in Fig. 1A, was the last to elute from the column, and this peak 5 fraction contained most of the heparanase activity. SDS-PAGE of this peak under non-reducing conditions revealed a high state of purity, with only two species of approximately 8 and 50 kDa, indicated by arrows in Fig. 1B. Both silver staining and immunostaining with anti-heparanase antibodies showed the presence of the 65-kDa protein in fraction 4, where the low heparanase activity would be consistent with the faint 50-kDa band in this fraction (Fig. 1, lane 4). The overall yield of the

The work described herein presents for the first time an understanding of the processing events leading to activation of heparanase and evidence that the active enzyme is a heterodimer. Others who have studied this enzyme have concluded that the 50-kDa protein alone is responsible for activity (11–13, 21, 22); no one has reported the presence of the 8-kDa peptide. A number of explanations might be considered here. The 8-kDa fragment stains less intensely on acrylamide gels than does the 50-kDa protein (Fig. 1B) and, unless one applies a gel system capable of resolving low molecular weight components, it is likely to go undetected, especially at the low levels of protein encountered with heparanase. Moreover, the 8-kDa peptide is NH<sub>2</sub> terminally blocked and refractive to Edman

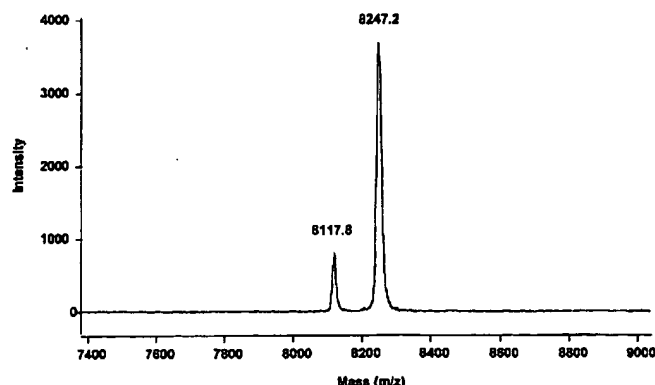


FIG. 4. MALDI-TOF analysis of the 8-kDa chain. The protein, purified by RP-HPLC (Fig. 2, peak 1) was concentrated in a Geloader<sup>TM</sup> pipette tip containing C18 packing, washed with water, and eluted onto a MALDI-TOF target with 80% acetonitrile in 0.1% TFA. Eluted sample was allowed to air dry to an approximate volume of 1  $\mu$ l and mixed with 1  $\mu$ l of sinapinic acid (10 mg/ml in 30% acetonitrile, 0.3% TFA). The sample was analyzed in linear delayed extraction mode and externally calibrated using insulin and thioredoxin. The peak of mass 8117.8 corresponds to a minor component extending from PCA<sup>36</sup> to Lys<sup>108</sup> (calculated mass = 8118.4 Da); the major product of mass 8247.2 Da represents the peptide extending from PCA<sup>36</sup> to Glu<sup>109</sup> (calculated mass = 8247.5 Da).

degradation, further complicating its identification in a mixture with the 50-kDa protein (12). Of course, it may be that the more extensive purification protocols reported earlier (21, 22) led to dissociation of the 50/8-kDa complex and that the 8-kDa chain is not required for activity. In our hands, however, the complex survived gel filtration at high ionic strength and in the presence of detergent, as well as two affinity purification steps. Furthermore, it is usually the case that processing of proteins from progenitor molecules does not produce the active entity as a stable complex with its activation peptide, unless that activity is dependent upon the complex itself. An apt example here is procaspase activation, where excision of a peptide bridging NH<sub>2</sub>-terminal and COOH-terminal polypeptides gives rise to the active 2-chain "heterodimeric" caspase (23). These chains remain in tight, non-covalent association, and both are obligate for function, while the bridge peptide is lost.

A final, and compelling argument for the importance of the 8-kDa chain derives from cloning and expression of heparanase done in other laboratories. Heparanase activity is easily demonstrated in mammalian cell hosts transfected with full-length cDNA corresponding to the 543-residue pre-proheparanase (11–13). However, expression of the truncated 50-kDa protein defined by Lys<sup>158</sup> to Ile<sup>543</sup> (12) failed to yield active enzyme, and this led to speculation that the region NH<sub>2</sub>-terminal to Lys<sup>158</sup> may play some functional role in catalysis. Indeed, our results would serve to define this region as the 8-kDa peptide.

Considering evidence presently in hand, we conclude that both the 8- and 50-kDa chains are required for enzyme activity, but we lack proof for this assertion. These protein components are easily separated under denaturing conditions on SDS-

PAGE (Fig. 1) or by RP-HPLC (Fig. 2). Unfortunately, solvent systems that promote protein unfolding are required for dissociation of the chains, and these conditions lead to irreversible loss of enzyme activity. Therefore, a reconstitution experiment with chains separated as shown in Fig. 2A has not been feasible and, at present, we lack definitive evidence for an obligate heterodimer.

Finally, an earlier report from this laboratory suggested that heparanase was a post-translationally modified form of a CXC chemokine, namely CTAPIII (7). We have not been able to confirm this observation, nor have others who have purified and characterized human heparanase (11–13, 21, 22). Yet, a recent paper by Rechter *et al.* (24) reported heparanase activity in a CTAPIII fusion protein with cellulose binding domain. Given this observation, the notion of a chemokine with heparanase activity remains a controversial issue.

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#### REFERENCES

1. Ruoslahti, E., and Yamaguchi, Y. (1991) *Cell* **64**, 867–869
2. Nakajima, M., Irimura, T., and Nicolson, G. L. (1998) *J. Cell. Biochem.* **36**, 157–167
3. Vlodavsky, I., Eldor, A., Haimovitz-Friedman, A., Matzner, Y., Ishai-Michaeli, R., Lider, O., Naparstek, Y., Cohen, I. R., and Fuks, Z. (1992) *Invasion Metastasis* **12**, 112–127
4. Thunberg, L., Backstrom, G., Wasteson, A., Robinson, C. N., Orgen, S., and Lindahl, U. (1982) *J. Biol. Chem.* **257**, 10278–10282
5. Abrahamsson, T., Brandt, U., Marklund, S. L., and Sjoquist, P. (1992) *Circ. Res.* **70**, 264–271
6. Tanaka, Y., Adams, D., and Shaw, S. (1993) *Immunol. Today* **14**, 111–115
7. Hoogewerf, A. J., Leone, J. W., Reardon, I. M., Howe, W. J., Asa, D., Heinrichson, R. L., and Ledbetter, S. R. (1995) *J. Biol. Chem.* **270**, 3268–3277
8. Kato, M., Wang, H., Kainulainen, V., Fitzgerald, M. L., Ledbetter, S., Ornitz, D. M., and Bernfield, M. (1998) *Nat. Med.* **4**, 691–697
9. Matzner, Y., Vlodavsky, I., Bar-Ner, M., Ishai-Michaeli, R., and Tauber, A. I. (1992) *J. Leuk. Biol.* **51**, 519–524
10. Lohse, D. L., and Linhardt, R. J. (1992) *J. Biol. Chem.* **267**, 24347–24355
11. Vlodavsky, I., Friedmann, Y., Elkin, M., Aingorn, H., Atzmon, R., Ishai-Michaeli, R., Bitan, M., Pappo, O., Peretz, T., Michal, I., Spector, L., and Pecker, I. (1999) *Nat. Med.* **5**, 793–802
12. Hulett, M. D., Freeman, C., Hamdorf, B. J., Baker, R. T., Harris, M. J., and Parish, C. R. (1999) *Nat. Med.* **5**, 803–809
13. Kussie, P. H., Hulmes, J. D., Ludwig, D. L., Patel, S., Navarro, E. C., Seddon, A. P., Giorgio, N. A., and Bohlen, P. (1999) *Biochem. Biophys. Res. Commun.* **261**, 183–187
14. Finkel, E. (1999) *Science* **285**, 33–34
15. Eccles, S. A. (1999) *Nat. Med.* **5**, 735–736
16. Ledbetter, S. R., Fisher, L. W., and Hassell, J. R. (1987) *Biochemistry* **26**, 988–995
17. Laemmli, U. K. (1970) *Nature* **227**, 680–685
18. Schagger, H., and Von Jagow, G. (1987) *Anal. Biochem.* **166**, 368–379
19. Switzer, R. C., III, Merrill, C. R., and Shifrin, S. (1979) *Anal. Biochem.* **98**, 231–237
20. Nielsen, H., Engelbrecht, J., Brunak, S., and von Heijne, G. (1997) *Protein Eng.* **10**, 1–6
21. Freeman, C., and Parish, C. R. (1998) *Biochem. J.* **330**, 1341–1350
22. Gonzalez-Stawinski, G. V., Parker, W., Holzknecht, Z. E., Huber, N. S., and Platt, J. L. (1999) *Biochim. Biophys. Acta* **1429**, 431–438
23. Cohen, G. M. (1997) *Biochem. J.* **326**, 1–16
24. Rechter, M., Lider, O., Cahalon, L., Baharav, E., Dekel, M., Seigel, D., Vlodavsky, I., Aingorn, H., Cohen, I. R., and Shoseyov, O. (1999) *Biochem. Biophys. Res. Commun.* **255**, 657–662